

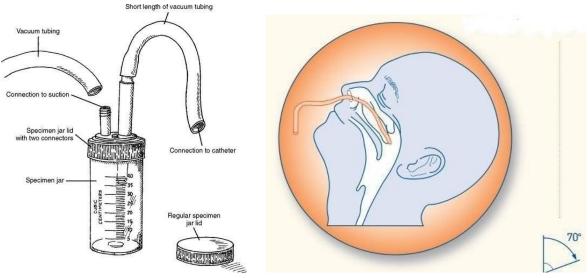
Nasal Aspirate:

This specimen source is acceptable for the following assays:

- Coronavirus SARS-CoV-2 by PCR (COVID-19)
- Coronavirus SARS-CoV-2 with Influenza AB RSV
- Respiratory Panel by PCR (without the SARS-CoV-2 target)
- Influenza AB RSV PCR

Materials:

- Portable suction pump
- Sterile suction catheter
- Mucus trap



Method:

- 1. Attach mucus trap to suction pump and catheter, leaving wrapper on suction catheter.
- 2. Turn on suction and adjust to suggested pressure (see Table 1).

Table 1		
Patient Age	Catheter Size	Suction Pressure
Premature Infant	6	80-100 mmHg
Infant	8	80-100 mmHg
Toddler/Preschooler	10	100-120 mmHg
School Age	12	100-120 mmHg
Adolescent/Adult	14	120-150 mmHg

3. Tilt the head back 70 degrees.



- 4. Without applying suction, insert catheter into nose, directed posteriorly toward the opening of the external ear.
 - To determine the length of catheter tubing needed, measure the distance from the tip of the nose to the external opening of the ear.
- 5. Apply suction; using a rotating movement, slowly withdraw catheter. Catheter should be in nasopharynx no longer than 10 seconds.
- 6. Rinse catheter with saline (if necessary), disconnect suction.
- 7. Remove the cap with the tubing and securely attach the **solid** lid.

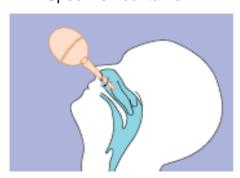
Nasal Wash:

This specimen source is acceptable for the following assays:

- Coronavirus SARS-CoV-2 by PCR (COVID-19)
- Coronavirus SARS-CoV-2 with Influenza AB RSV
- Respiratory Panel by PCR (without the SARS-CoV-2 target)
- Influenza AB RSV PCR

Materials:

- Saline
- 1-2 oz. Tapered rubber bulb
- 3-5 mL syringe and 18 20-gauge tubing (if using the syringe method).
- Specimen container





Bulb Method:

- 1. Suction 3-5 mL saline into a new sterile bulb.
- 2. Insert bulb into one nostril until nostril is occluded.
- 3. Instill saline into nostril with one squeeze of the bulb.
- 4. Immediately release bulb to collect recoverable specimen.
- 5. Empty bulb into suitable dry, sterile specimen container or test tube with cap.

Syringe Method:



- 1. Same process as the bulb method by injecting saline and quickly aspirate back into syringe and cap the syringe.
- 2. Empty syringe into suitable dry, sterile specimen container or test tube with cap.

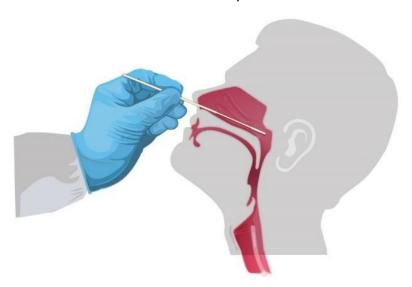
Nasopharyngeal Swabs:

This specimen source is acceptable for the following assays:

- Coronavirus SARS-CoV-2 by PCR (COVID-19)
- Coronavirus SARS-CoV-2 with Influenza AB RSV
- Respiratory Panel by PCR with SARS-CoV-2
- Respiratory Panel by PCR (without the SARS-CoV-2 target)
- Influenza AB RSV PCR

Materials:

- Flocked or polyester-tipped nasopharyngeal swabs with a flexible shaft (wire or plastic)
- 1.5-3 mL viral or universal transport medium



Method:

- 1. Tilt patient's head back 70 degrees.
- Gently and slowly insert the swab through the nostril parallel to the palate (not upwards). Do this until resistance is encountered or the distance is equivalent to that from the ear to the nostril of the patient. This indicates contact with the nasopharynx.



- 3. Gently rub and roll the swab. Leave the swab in place for several seconds to absorb secretions.
- 4. Slowly remove swab while rotating it.
 - NOTE: If a deviated septum or blockage create difficulty in obtaining the specimen from one nostril, use the same swab to collect the specimen from the other nostril.

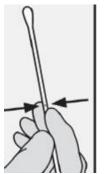
Nasal Swab:

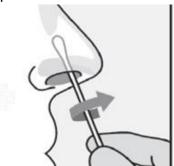
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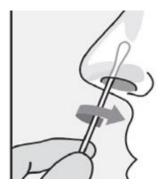
Coronavirus SARS-CoV-2 by PCR (COVID-19)

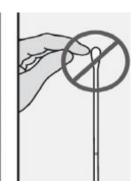
Materials:

- Flocked or polyester-tipped nasal swabs with plastic shaft.
- Viral transport medium.









Method:

- 1. Hold the swab with the score line above your hand, do not pre-wet swab in media.
- 2. Insert swab 1-2 cm into one of the anterior nares.
- 3. Rotate swab against nasal mucosa for 3 seconds and withdraw.
- 4. Repeat in the other anterior nares using the same swab.
- 5. Place swab **immediately** into sterile tube containing 1.5-3 mL viral or universal transport medium.

SARS Antigen FIA:

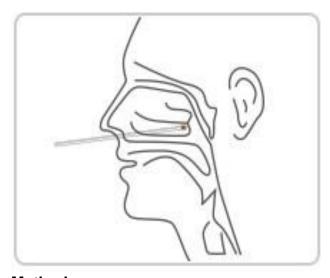
Materials:

- Nasal Swab only. The Puritan Sterile Foam tipped Applicators (pictured) are the only acceptable swabs.
- Sterile blue capped tube or sterile urine cup.









Method:



- 1. Prior to collecting the nasal swab, the patient should be instructed to blow their nose.
- 2. Carefully insert the swab into the nostril presenting with the most secretion under visible inspection. Using a gentle rotation, push the swab until resistance is met at the level of the turbinate (less than 1 inch into the nostril).
- 3. Rotate the swab several times against the nasal wall then remove it from the nostril.
- 4. Place swab in sterile tube or cup.

NOTE: Swab may need to be cut to fit in sterile container. Specimens should be tested as soon as possible.